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Extracellular nicotinamide phosphoribosyltransferase (NAMPT) promotes M2 macrophage polarization in chronic lymphocytic leukemia

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Abstract

Nicotinamide phosphoribosyltransferase (NAMPT) is the rate-limiting enzyme in nicotinamide adenine dinucleotide biosynthesis. In the extracellular compartment, it exhibits cytokine-/adipokinelike properties, suggesting that it stands at the crossroad between metabolism and inflammation. Here we show that both intracellular and extracellular NAMPT levels are increased in cells and plasma of chronic lymphocytic leukemia (CLL) patients. The extracellular form (eNAMPT) is produced by CLL lymphocytes upon B-cell receptor, Toll-like receptor, and nuclear factor κ B (NF- κ B) signaling pathway activation. eNAMPT is important for differentiation of resting monocytes, polarizing them toward tumor-supporting M2 macrophages. These cells express high levels of CD163, CD206, and indoleamine 2,3-dioxygenase and secrete immunosuppressive (interleukin [IL] 10, CC chemokine ligand 18) and tumor-promoting (IL-6, IL-8) cytokines. NAMPT-primed M2 macrophages activate extracellular-regulated kinase 1/2, signal transducer and activator of transcription 3, and NF- κ B signaling; promote leukemic cell survival; and reduce T-cell responses. These effects are independent of the enzymatic activity of NAMPT, as inferred from the use of an enzymatically inactive mutant. Overall, these results reveal that eNAMPT is a critical element in the induction of an immunosuppressive and tumor-promoting microenvironment of CLL.

Introduction

Besides being the first line of defense against pathogens, macrophages orchestrate tissue plasticity and homeostasis. They are classified into classically activated (M1) or alternatively activated (M2) macrophages, reflecting a different functional role.¹ In cancer tissues, macrophages tend to be of the M2 phenotype, acquired and maintained through multiple interactions with tumor cells.² Evidence indicates that these macrophages enhance tumor progression, mainly through the secretion of chemokines/cytokines that sustain neoplastic cell proliferation and suppress immune responses.^{3,4}

Chronic lymphocytic leukemia (CLL) is a disease of mature B cells, which rely on the host environment for progression.⁵⁻⁷ Tumor-host interactions occur predominantly in protected niches in the lymph nodes (LNs) and in the bone marrow, known as proliferation centers.^{8,9} Within these areas, CLL cells are in contact with a population of CD68⁺ elements, resembling tumor-associated macrophages.¹⁰⁻¹³ They may be also differentiated in vitro by coculturing peripheral blood monocytes with CLL cells. These so-called nurselike cells (NLCs) protect leukemic cells from apoptosis through multiple interactions regulated by soluble or cell-surface-anchored molecules.^{14,15} Leukemic cells play an essential role in driving NLC differentiation, as inferred from the lack of differentiation when monocytes from CLL patients are cultured with normal B lymphocytes.¹⁶ However, the signals and factors that regulate NLC differentiation are incompletely understood.

The enzyme nicotinamide phosphoribosyltransferase (NAMPT) was first identified in the supernatants of activated lymphocytes during the search for novel cytokinelike proteins.¹⁷ A few years later, NAMPT, dubbed visfatin, was recognized as a novel adipokine exerting insulinlike properties.¹⁸ The unexpected finding that the same protein possessed significant homology with a bacterial enzyme, termed NadV, turned NAMPT into a unique enzyme-cytokine molecule.^{19,20} NAMPT utilizes 5-phosphoribosyl-1-pyrophosphate (PRPP) and nicotinamide to generate nicotinamide mononucleotide (NMN), which is subsequently adenylated to nicotinamide adenine dinucleotide (NAD) by the enzyme NMN adenylyltransferase. The NAMPT-catalyzed reaction is considered the rate-limiting step in NAD biosynthesis

from nicotinamide.^{20,21} The cytokinelike functions of NAMPT prevail in the extracellular environment, where the enzymatic activity seems to be dispensable.^{22,23} Extracellular NAMPT levels are increased in various metabolic and inflammatory diseases,^{24,25} as well as in tumors,^{26–29} rendering this pleiotropic molecule a novel player in tumor/host cross-talk.

This work shows that NAMPT levels are increased in CLL cells and that extracellular NAMPT (eNAMPT) production is induced upon activation of the leukemic cell. In the tumor microenvironment, eNAMPT is an important element in inducing monocyte polarization to M2 macrophages secreting tumor-promoting cytokines and inhibiting T-cell responses. Lastly, drugs that block CLL cell activation or that restore immune competence decrease eNAMPT production.

Methods

Patient and healthy donor (HD) samples

The study was approved by the Human Genetics Foundation Ethical Committee. Blood samples were obtained in accordance with Institutional Guidelines and the Declaration of Helsinki. Patient characteristics are reported in supplemental Table 1 (available on the *Blood* Web site). Blood samples of HDs were obtained through the local blood bank.

Purified B lymphocytes were prepared as described³⁰ and cultured as detailed in supplemental Methods.

Antibodies and reagents

The full list of antibodies and reagents is provided in supplemental Methods.

Purification of monocytes and NLC generation

Circulating monocytes were isolated by cell sorting using a fluorescence-activated cell sorter (FACS) ARIA III sorter (BD Biosciences, Milan, Italy). NLCs were generated as described.³¹ When indicated, recombinant NAMPT (rNAMPT; 200 ng/mL, Adipogen, San Diego, CA), blocking anti-NAMPT (50 µg/mL) polyclonal antibody (pAb),^{32,33} or lenalidomide (0.5 µM) were added to the cultures.

NAMPT activity in lysates or plasma

NAMPT activity was determined by a novel, multicoupled fluorometric assay, developed to measure NAMPT activity in cell crude extracts and biological fluids.³⁴ Full details are provided in supplemental Methods.

NAD and NMN determination

NAD was quantitated by the fluorometric cycling assay.³⁵ NMN was measured upon derivatization with acetophenone and spectrofluorometric high-performance liquid chromatography analysis,³⁶ with modifications specified in supplemental Methods.

eNAMPT quantification by enzyme-linked immunosorbent assay (ELISA)

eNAMPT concentrations were determined using human NAMPT ELISA kit (Adipogen).

Immunocytochemistry

Cell morphology and numbers were studied by Giemsa staining.³⁷ For immunocytochemistry, coverslips were stained as detailed in supplemental Methods.

Immunohistochemistry

Formalin-fixed, paraffin-embedded sections of CLL (n = 9) or reactive (n = 3) LNs were stained as described³⁸ and analyzed by light microscopy as detailed in supplemental Methods.

Confocal microscopy

Slides were analyzed using a TCS SP5 laser scanning confocal microscope; images were acquired with LAS AF software (both from Leica Microsystems, Milan, Italy). Pixel intensity was calculated using the ImageJ software (<http://rsbweb.nih.gov.offcampus.dam.unito.it/ij/>).

Phagocytosis assay

NLCs were incubated (15 minutes, 37°C) with fluorescein isothiocyanate (FITC)-dextran (1 mg/mL in phosphate-buffered saline [PBS] + 5% fetal calf serum [FCS]) to allow internalization. Where indicated, NLCs were pretreated with neutralizing monoclonal antibody to CD206 (60 minutes, 37°C).³⁹ Coverslips were stained⁴⁰ and analyzed by confocal microscopy.

FACS analyses

Data were acquired using a FACSCanto II cytofluorometer (BD Biosciences) and processed with DIVA-v7 (BD Biosciences) and FlowJo-v9.01 softwares (TreeStar, Ashland, OR).

Immunoprecipitation and western blot

Supernatants and CLL plasma samples were concentrated using Microcon 30k (Merck-Millipore, Vimodrone, Italy). Albumin was removed using Albumin Depletion Kit (Pierce-Thermo-Scientific, Rockford, IL). Anti-NAMPT monoclonal antibody (Adipogen) was employed for immunoprecipitation (Protein-G-Mag Sepharose; GE Healthcare, Milan, Italy).

Whole cell lysates⁴¹ were resolved by sodium dodecyl sulfate–polyacrylamide gel electrophoresis and transferred to nitrocellulose membranes (Bio-Rad, Hercules, CA). rNAMPT (200 ng, Adipogen) was used as reference control. The shift in molecular weight between endogenous and rNAMPT is because of the FLAG (DYKDDDDK sequence peptide) tag.

Images were acquired using the ImageQuant Las4000 gel imager (GE Healthcare).

RNA extraction and quantitative real-time polymerase chain reaction (qRT-PCR)

RNA was extracted using RNeasy Plus Mini kit (Qiagen, Milan, Italy) and converted to complementary DNA using the High Capacity cDNA Reverse Transcription kit (Life Technologies, Milan, Italy). qRT-PCR was performed using the 7900 HT Fast Real Time PCR system (SDS2.3 software) using the TaqMan assays (Life Technologies) listed in supplemental Methods. Relative gene expression was calculated as described.⁴²

Cell viability, proliferation, and chemotaxis assays

Cell viability was measured using the Annexin-V Apoptosis Kit (Valter Occhiena, Turin, Italy), and proliferation using carboxyfluorescein diacetate succinimidyl ester (CFSE; Life Technologies). Chemotaxis experiments were performed using classical Boyden chamber assays.³⁰

Cytokine/chemokine measurement

Interleukin (IL) 6, IL-8, IL-10, and CC chemokine ligand (CCL) 3 concentrations were determined using the Bio-Plex/Luminex technology (<http://www.bioclarma.com>). CCL18 was determined using a specific ELISA assay (R&D Systems, Milan, Italy).

T-cell proliferation

Coculture of NLCs with autologous peripheral blood mononuclear cells (PBMCs) was performed after CFSE labeling of preactivated PBMCs with anti-CD3 (2 µg/mL) and IL-2 (15 IU/mL) for 3 to 5 days with or without rNAMPT.⁴³ T-cell proliferation was analyzed by flow cytometry, after gating on CD3⁺ lymphocytes.

Statistical analysis of data

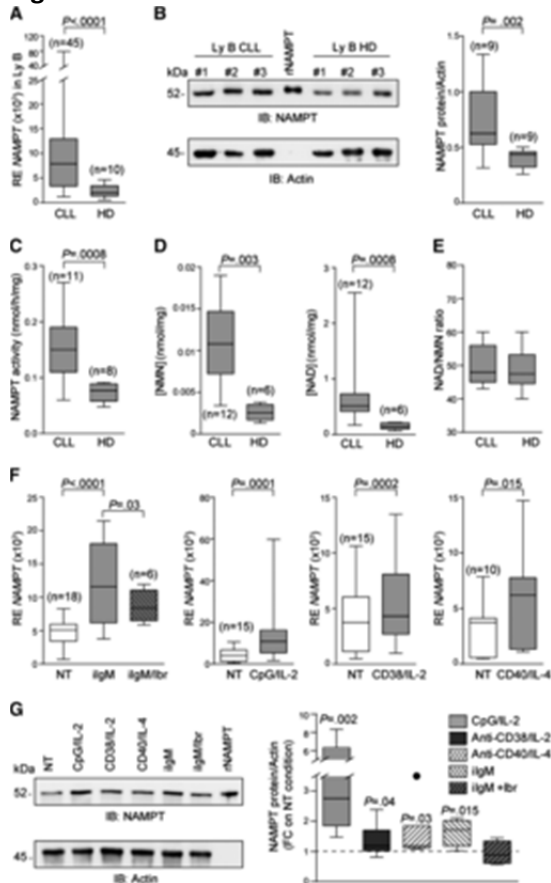
Statistical analyses were performed with GraphPad version 6.0 (GraphPad Software Inc., La Jolla, CA). Continuous variables were compared by Mann-Whitney *U* (unpaired data) or Wilcoxon signed rank (paired data) tests. Correlation between continuous variables was assessed using Pearson's coefficient.

Results

Intracellular NAMPT (iNAMPT) is overexpressed in CLL cells and is enzymatically active

NAMPT messenger RNA (mRNA) and protein expression in lysates of CLL and normal B lymphocytes were comparatively investigated by qRT-PCR and western blot. *NAMPT* mRNA levels in purified CLL cells were significantly higher than those of normal B lymphocytes obtained from the peripheral blood of age- and sex-matched donors (Figure 1A). Analysis of protein levels in whole cell lysates confirmed higher levels of iNAMPT in purified CLL compared with normal circulating B cells (Figure 1B). Accordingly, the activity of the enzyme was higher in CLL than in normal B lymphocytes (Figure 1C). In keeping with the *NAMPT* activity profile, CLL cells also contained higher NMN and NAD levels than normal B lymphocytes (Figure 1D). The NAD/NMN ratio was constant and similar in all cells tested (Figure 1E), confirming that the enzyme converting NMN to NAD (ie, NMN adenylyltransferase) is not the rate-limiting enzyme in the nicotinamide-NAD pathway.⁴⁴

Figure 1



An enzymatically active NAMPT is overexpressed by CLL cells. (A) Box plot showing expression levels of *NAMPT* mRNA in B lymphocytes from CLL patients (n = 45) or HDs (n = 10). (B) Western blot analysis of NAMPT protein expression in CLL (n = 9) or HD (n = 9) B lymphocytes. A FLAG-tagged rNAMPT was used as internal control. (C-E) Box plots representing NAMPT activity expressed as nmol/hour/mg of protein (C), NMN and NAD intracellular concentration (nmol/mg) (D), and NAD/NMN ratio (E) in B lymphocytes from CLL patients and HDs. (F) qRT-PCR analysis showing expression of *NAMPT* mRNA in purified CLL lymphocytes cultured with the indicated stimuli (24 hours). When indicated, cells were pretreated with ibrutinib (10 μ M, 30 minutes). (G) Purified CLL cells activated as indicated in panel F were lysed, and iNAMPT expression levels determined by western blot. Cumulative results (n = 8) are shown in the box plot.

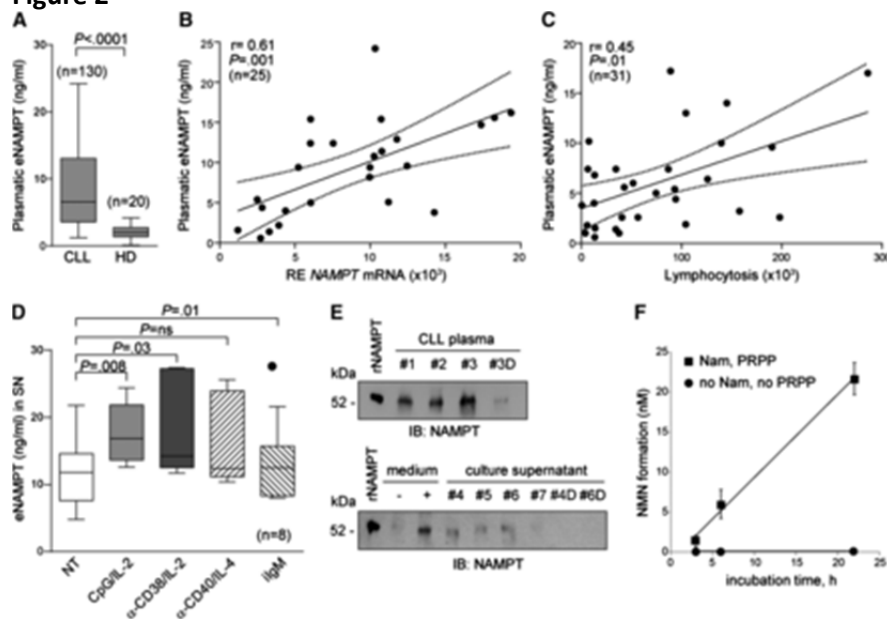
When CLL signaling pathways were activated, *NAMPT* mRNA was significantly upregulated. The increase in *NAMPT* mRNA could be highlighted after engagement of B-cell receptor (BCR), Toll-like receptor 9 (TLR9), CD38, and CD40 (24 hours, Figure 1F). These signals are known to induce CLL cell activation and *CCL3* gene transcription,^{45,46} as confirmed here (supplemental Figure 1A). Higher levels of *NAMPT* mRNA were paralleled by increased intracellular protein (Figure 1G). Treatment of activated CLL cells with ibrutinib, a selective bruton tyrosine kinase (BTK) inhibitor, abrogated BCR-, TLR9-, and CD38-induced *NAMPT* mRNA and protein upregulation (Figure 1F-G and supplemental Figure 1B-C), confirming that BTK is a critical intermediate in these signaling pathways^{47,48} and linking this kinase with *NAMPT* transcription. In the conditions used⁴⁹ (10 μ M dose, 24 hours), we detected complete inhibition of BTK phosphorylation, with limited cytotoxicity (supplemental Figure 1D-E).

NAMPT is present in plasma and can be secreted by activated CLL cells

We then asked whether NAMPT protein could also be found in the extracellular environment (eNAMPT). By using an ELISA assay, eNAMPT was detected in the plasma of CLL patients (n = 130) at significantly higher concentrations than in HDs of a comparable age (Figure 2A). The finding of a positive correlation between *NAMPT* mRNA expression in purified CLL cells and eNAMPT levels in plasma (Figure 2B) suggested that leukemic lymphocytes may contribute to the production of eNAMPT. Consistently, a positive correlation

between plasmatic NAMPT and absolute lymphocytosis was observed in CLL patients with comparable percentages of circulating monocytes, known contributors to NAMPT secretion (Figure 2C).⁵⁰

Figure 2



eNAMPT is present in CLL plasma and can be produced by activated CLL cells. (A) Box plot showing eNAMPT concentrations measured with a quantitative ELISA assay performed on plasma samples from CLL patients ($n = 130$) or HDs of a comparable age ($n = 20$). (B) Regression line showing a positive correlation between NAMPT mRNA levels (x-axis) and plasmatic eNAMPT (y-axis) in 25 CLL patients. Pearson coefficient (r) and the corresponding P value are noted. (C) Regression line showing a positive correlation between lymphocytosis (x-axis) and plasmatic eNAMPT (y-axis) in 31 CLL patients. Pearson coefficient (r) and the corresponding P value are noted. (D) ELISA assay showing eNAMPT concentrations in supernatants (SN) from cultures of purified CLL cells cultured with the indicated stimuli ($n = 8$, 24 hours). (E) The presence of eNAMPT was confirmed by western blot performed on concentrated ($\times 10$) albumin-depleted CLL plasma samples (#1-#3) and culture supernatants (#4-#7) from different CLL patients with high (>20 ng/mL, #4-#6) or low (<5 ng/mL, #7) eNAMPT levels, as detected by ELISA assay. The condition (–) and (+) medium indicates concentrated ($\times 10$) albumin-depleted RPMI +10% FCS alone (–) or with 20 ng of rNAMPT (+). #3D, #4D, #6D indicate NAMPT-depleted fractions by immunoprecipitation from #3 CLL plasma, or from supernatants #4 and #6. rNAMPT was loaded as control. (F) Time course of eNAMPT activity determined in the plasma of a prototype CLL patient, as described in supplemental Methods.

Culture of CLL cells for 24 hours was followed by the appearance of eNAMPT in spent media (Figure 2D), independently of the number of cells undergoing apoptosis or proliferation, evincing an active rather than a passive mechanism (not shown). Furthermore, after activation of purified CLL lymphocytes through the BCR, TLR9, CD38, or CD40 for 24 hours, eNAMPT levels were invariably increased (Figure 2D). No significant modification in the total number of cells was apparent at this time point. The nature of the protein present in CLL plasma or culture supernatants (Figure 2E and supplemental Figure 2A) was confirmed by western blot, showing a protein compatible with NAMPT selectively in eNAMPT-rich samples. Furthermore, deprivation of eNAMPT by immunoprecipitation was followed by the disappearance of the western blot band (Figure 2E).

These results suggest that eNAMPT could derive, at least partly, from leukemic cells. The generation of eNAMPT from CLL cells is apparent under resting conditions and increases upon activation, arguing in favor of a role for this cytokine/enzyme in the CLL microenvironment.

The critical issue in the field is whether eNAMPT is enzymatically active and whether this activity is relevant in the extracellular compartment.^{22,51,52} Using an ad hoc devised assay,³⁴ we detected NMN production when CLL plasma was incubated with nicotinamide and PRPP (Figure 2F), with an activity of 1.7 ± 0.1 pmol NMN/hour per mL plasma. However, in the absence of added nicotinamide and PRPP, NAMPT activity was

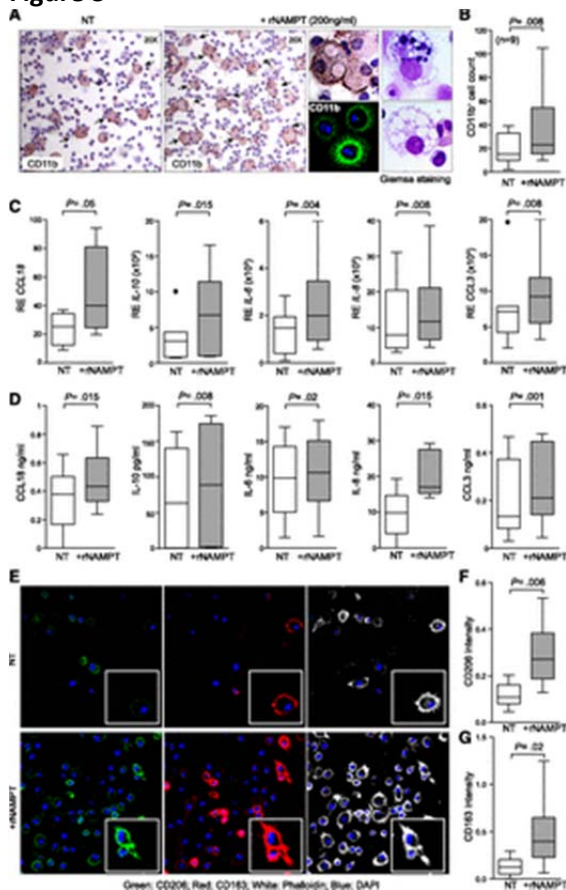
undetectable, in line with previous studies.⁵³ Accordingly, NMN, the product of the reaction, was undetectable in both plasma and media (supplemental Figure 2B).

Treatment of CLL monocytes with NAMPT induces M2 macrophage differentiation

Next we analyzed the functional role of eNAMPT in the CLL microenvironment. Exposure of purified CLL cells to rNAMPT did not result in increased proliferation or chemotaxis, but it significantly enhanced survival of the leukemic cells (supplemental Figure 3A-C). The protective effect may be explained by de novo transcription of *TNF- α* (supplemental Figure 3D), which is involved in antiapoptotic effects through the activation of B cell activating factor and a proliferation inducing ligand.^{54,55} Conversely, the increased transcription of *TGF- β* and of *CCL3* (supplemental Figure 3E-F) in response to rNAMPT argued in favor of a “microenvironmental effect” of eNAMPT. This later hypothesis is in agreement with previous data implicating the molecule in the differentiation and activation of monocytes.⁵⁶⁻⁵⁸

For this reason, we further investigated whether eNAMPT produced by activated leukemic cells modifies the myeloid component. After 5 days of exposure to rNAMPT, PBMCs from normal donors showed a significant increase in CD11b⁺ macrophages (supplemental Figure 4A-C). This finding was confirmed using PBMCs of CLL patients. Macrophage increases were substantiated by Giemsa staining (highlighting typical intracellular vacuoles and granules) and by CD11b and CD68 staining (Figure 3A-B and supplemental Figure 5A-D).

Figure 3



rNAMPT induces monocytes to differentiate into macrophages with M2 features. (A) PBMC preparations from CLL patients were plated in complete medium with or without rNAMPT (200 ng/mL). After 5 days, cells were fixed and stained for CD11b. Original magnification $\times 20$. Middle panel represents a zoomed area of the same sample. CD11b staining was also confirmed by immunofluorescence. The panels on the right show Giemsa staining to evaluate cellular morphology. (B) Cumulative data showing the number of CD11b⁺ cells in at least 4 $\times 20$ fields of 9 different samples. (C-D) Box plots reporting the mRNA (C) or the protein (D) expression levels of CCL18, IL-10, IL-6, IL-8 and CCL3, in monocytes sorted from leukemic PBMCs (n = 9) cultured with or without

rNAMPT (24 hours). (E) Confocal microscopy analysis of CD163 (red) and CD206 (green) expression in sorted CLL monocytes differentiated for 5 days with or without rNAMPT. Nuclei were counterstained with 4,6-diamidino-2-phenylindole (DAPI; blue); actin filaments were visualized using phalloidin (white). Original magnification ×63. (F-G) Cumulative analysis of CD206 (F) and CD163 (G) pixel intensity (arbitrary units, a.u.), scoring at least 10 different cells for 3 different samples.

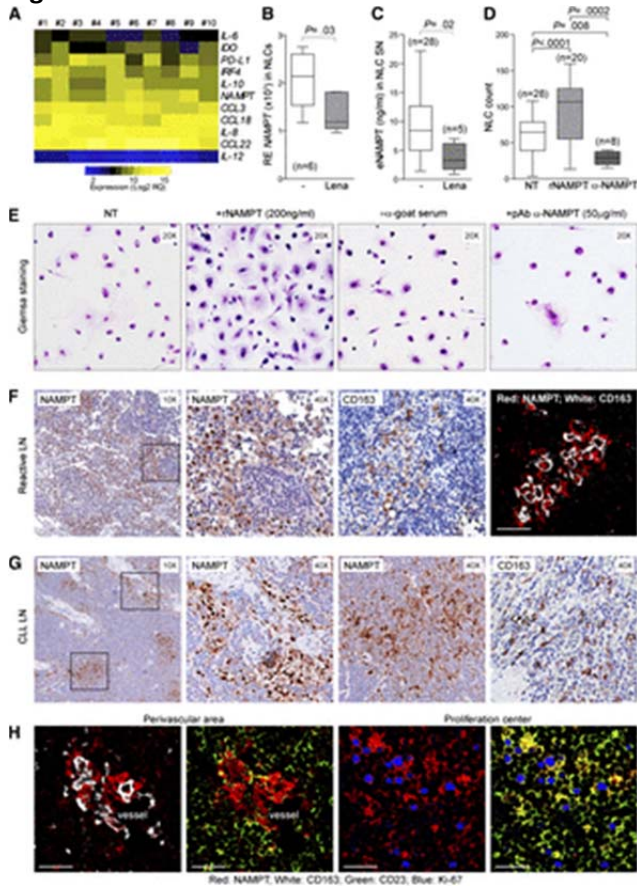
These results suggest the existence of a paracrine circuit, where activated CLL cells produce eNAMPT, that in turn recruits monocytes through CCL3 secretion⁵⁹ and induces macrophage differentiation. The use of purified monocyte preparations confirmed that eNAMPT acts directly on this cell population (supplemental Figure 5E).

The monocyte subset of CLL patients showed constitutive M2 skewing, based on high interferon regulatory factor 4 (*IRF4*) transcription factor and low *IL-12* levels, considered M2 and M1 markers, respectively (supplemental Figure 6A). Surface expression of the scavenger receptor CD163 and of the mannose receptor CD206, considered M2 markers, strengthened this finding (supplemental Figure 6B).^{60,61} Treatment of purified CLL monocytes with rNAMPT further enhanced M2 features, as inferred on the basis of (1) a transcriptional profile, which showed induction of *CCL18*, *IL-10*, *IL-6*, *IL-8*, and *CCL3* genes (Figure 3C); (2) evidence of increased concentrations of the same cytokines/chemokines in the supernatants of rNAMPT-treated monocytes (Figure 3D); and (3) increased expression of CD163 and CD206 (Figure 3E-G). Similar effects following rNAMPT treatment were observed when using monocytes from HDs (supplemental Figure 7A-B), suggesting that both HD and CLL monocytes can respond to rNAMPT.

NLCs express high levels of NAMPT

Monocytes from CLL patients spontaneously differentiate in vitro into NLCs, large, multinucleated, adherent cells that protect the leukemic clone from apoptosis.^{14,16} NLCs are also present in lymphoid tissues where they presumably deliver prosurvival signals to CLL cells.⁶² In agreement with evidence suggesting that NLCs are closely related to tumor-associated macrophages,^{11,12,43} they expressed high levels of *IRF4* and of the tryptophan-metabolizing enzyme indoleamine 2,3-dioxygenase (*IDO*; Figure 4A). Their chemokine/cytokine profile was typical of M2 macrophages, including *IL-10*, *IL-6*, *IL-8*, *CCL18*, *CCL22*, and *CCL3*. *PD-L1*, the ligand for the inhibitory programmed cell death-1 (PD-1) receptor, was also highly expressed. Conversely, *IL-12* mRNA, which distinguishes M1 macrophages, was undetectable (Figure 4A), as was expression of the costimulatory molecule CD80 (supplemental Figure 8A).

Figure 4



NAMPT is expressed by NLCs and by myeloid elements in CLL LNs. (A) Heat map showing gene expression profiling performed on conventionally differentiated NLCs ($n = 10$) highlighted elevated levels of genes classically associated with M2 macrophages, including *NAMPT*. Expression values are represented as Log2 of relative quantification (RQ) calculated as relative expression on *ACTB* housekeeping gene. (B-C) Box plot showing *NAMPT* mRNA expression levels (B) and eNAMPT soluble levels (C) in NLCs from CLL patients obtained using conventional culture conditions ($n = 28$) or differentiated in the presence of lenalidomide ($n = 5$). (D-E) NLC numbers (D) and morphology assessed by Giemsa staining (E) were determined in CLL samples differentiated with or without addition of rNAMPT. A blocking pAb anti-NAMPT or control preimmune goat serum were added to conventional NLC cultures to inhibit constitutive eNAMPT. Original magnification $\times 20$ in panel E. (F-G) Immunohistochemical analysis of NAMPT expression in reactive (F) or CLL (G) LN. CD163 was used to detect macrophages in both normal tissues and CLL LN. Images at $\times 4$ and $\times 20$ original magnifications. The immunofluorescence image in panel F shows complete overlap between NAMPT (red) and CD163 (white) staining in reactive LN (magnification $\times 63$). (H) Immunofluorescence images showing partial overlap between NAMPT staining (red) and CD163 (white) in CLL LN samples. Within the proliferation center, NAMPT shows partial colocalization with CD23⁺/Ki-67⁺ CLL lymphocytes. Original magnification $\times 63$.

In NLC samples, *NAMPT* mRNA was invariably elevated (Figure 4A-B). Moreover, conditioned media from NLC cultures contained high amounts of eNAMPT (Figure 4C). Differentiation of NLCs from CLL PBMC preparations was significantly increased when rNAMPT was added to the cultures during the differentiation process (Figure 4D). This conclusion was based on (1) morphologic parameters (Figure 4E) and (2) expression of lineage-specific markers such as CD11b and CD68 (supplemental Figure 8B-D), suggesting that NAMPT is important for full NLC differentiation. Confirmation was obtained by deriving NLCs in the presence of a blocking anti-NAMPT goat pAb,³³ which significantly decreased the total number of differentiated cells (Figure 4D-E). Furthermore, treatment with the immunomodulatory drug lenalidomide, which partially corrects the M2 phenotype,⁶³ robustly decreased *NAMPT* mRNA levels and in eNAMPT concentrations in spent media (Figure 4B-C), indicating that NAMPT is part of the M2 signature. As expected, *IRF4* expression was significantly downregulated (supplemental Figure 8E).⁶⁴

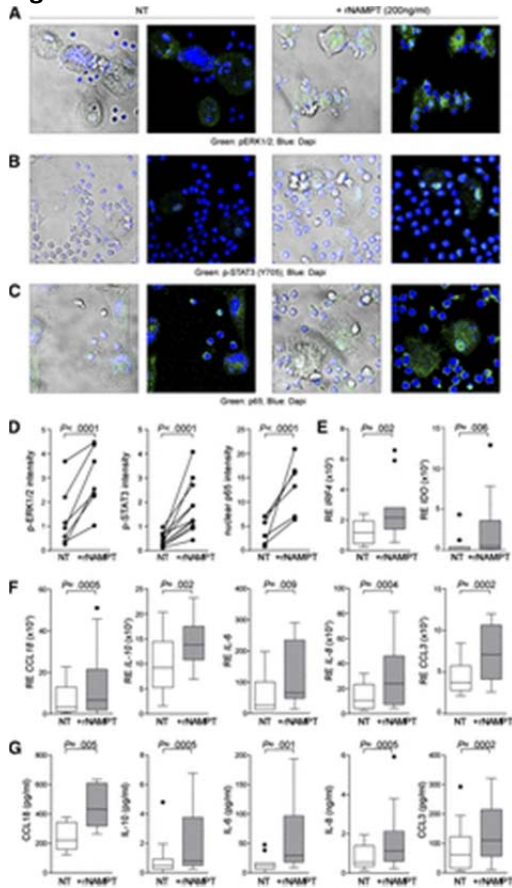
NAMPT is expressed in the proliferation centers of CLL LN

In reactive LN samples NAMPT was expressed by residential macrophages (CD11b⁺/CD68⁺/CD163⁺; Figure 4F and supplemental Figure 9A), as inferred both from morphologic parameters and from colocalization between NAMPT and CD163 fluorescent signals (Figure 4F). Accordingly, by qRT-PCR myeloid cells in the peripheral blood express the highest *NAMPT* levels, both in HD and CLL patients (supplemental Figure 8F). Residential CD68⁺/CD163⁺ macrophages clearly stained positive for NAMPT also in CLL LN samples (Figure 4G and supplemental Figure 9B). However, NAMPT⁺ elements were also present in the paler areas classically associated with proliferation centers (Figure 4G). NAMPT⁺ cells were a mixture of CD163⁺ and CD23⁺ elements, confirming that the leukemic clone can express NAMPT at high levels, as observed in vitro (Figure 4H and supplemental Figure 9C-D). In general, NAMPT⁺ cells overlapped or were in contact with Ki-67⁺ cells (Figure 4H and supplemental Figure 9E-F), consistent with our data indicating upregulation of NAMPT in proliferating CLL cells in culture.

Treatment of NLCs with rNAMPT induces chemokine secretion through the activation of NF-κB and STAT3

The final phase of the study examined the effects induced by rNAMPT on NLCs. Exposure of conventionally differentiated NLCs to rNAMPT induced a rapid and prominent activation of intracellular signaling. Phosphorylation of extracellular regulated kinase 1/2 (ERK1/2) started a few minutes after rNAMPT exposure, peaked at 20 to 30 minutes, and slowly decreased (Figure 5A,D). Nuclear translocation of phospho-signal transducer and activator of transcription 3 (p-STAT3) and nuclear factor κB (NF-κB) component p65 was also observed with comparable kinetics (Figure 5B-D), consistent with what was reported for murine macrophages²² and a human monocytic cell line.⁶⁵ Pretreatment with a blocking anti-NAMPT pAb completely inhibited signal transduction (supplemental Figure 10A-D). Furthermore, exposure of NLCs to rNAMPT for 24 hours was followed by enhanced transcription of *IRF4*, *IDO*, *CCL18*, *IL-10*, *IL-6*, *IL-8*, and *CCL3* (Figure 5E-F), the same transcripts that were previously found in rNAMPT-treated monocytes (Figure 3C-D). The corresponding cytokines and chemokines were present in media of rNAMPT-treated NLC preparations (Figure 5G). rNAMPT treatment in the presence of specific inhibitors of both NF-κB and STAT3 failed to modulate cytokine transcription, clearly indicating that activation of both pathways is required to drive gene expression in response to NAMPT (supplemental Figure 10E-I).

Figure 5



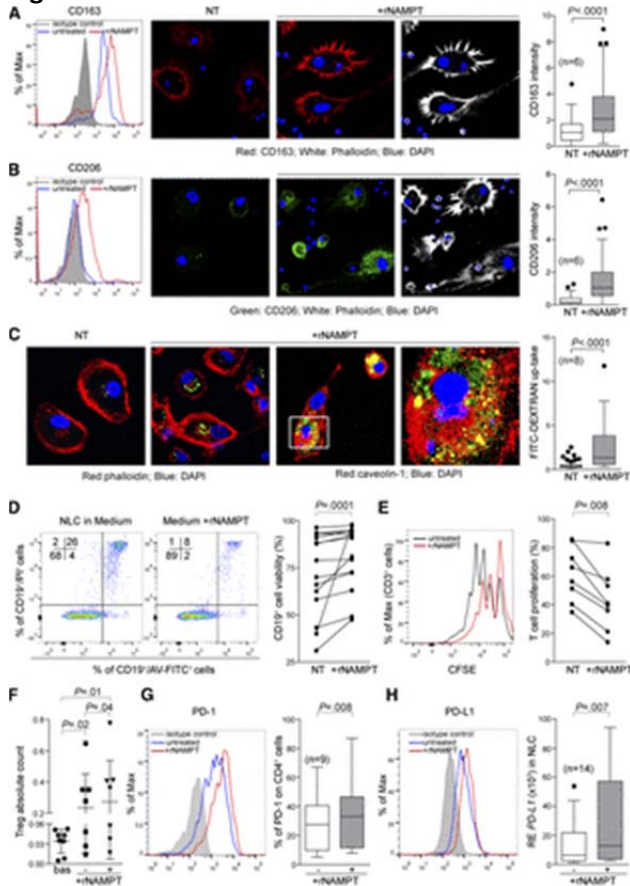
Exposure of NLCs to rNAMPT activates signal transduction and expression of a panel of genes specifying the M2 phenotype. NLCs were exposed to rNAMPT (30 minutes, 37°C) before fixing, permeabilizing, and staining for phospho-ERK1/2 (A), p-STAT3 (B), and the p65 subunit of the NF- κ B complex (C). DAPI (blue) was used to counterstain. Original magnification $\times 63$. (D) Graphs show cumulative analyses plotting the mean values of all the fluorescence measurements for each independent experiment. At least 4 randomly chosen fields from different samples were analyzed after defining a region of interest. (E-F) Conventionally differentiated NLCs were treated with rNAMPT for 24 hours before RNA extraction and qRT-PCR analysis of the expression of *IRF4* and *IDO* (E) or for a panel of chemokines/cytokines defining the M2 phenotype (F). (G) Increased expression of immunosuppressive chemokines/cytokines in cultured supernatants ($n = 14$) was confirmed at the protein level.

These results suggest that eNAMPT enhances M2 features in fully differentiated NLCs as well.

rNAMPT enhances the immunosuppressive features of NLCs

rNAMPT-treated NLCs were characterized by increased expression of the M2 macrophage markers CD163 (Figure 6A) and CD206 (Figure 6B). They were also characterized by increased phagocytosis of FITC-conjugated dextran particles (Figure 6C), which was however independent of CD206, as inferred by using a neutralizing anti-CD206 antibody (supplemental Figure 11).³⁹

Figure 6



Exposure of NLCs to rNAMPT enhances their immunosuppressive function. NLCs ($n = 6$) were generated in the presence of rNAMPT before assessing expression of CD163 (A) and CD206 (B) by flow cytometry and confocal microscopy. Box plots show cumulative analyses of pixel intensity (a.u.) in at least 4 fields for the different samples. (C) Conventionally obtained NLCs ($n = 8$) were incubated (15 minutes, 37°C) with FITC-dextran (1 mg/mL in PBS + 5% FCS) with or without rNAMPT. Phagocytosis was confirmed by costaining with caveolin-1 (in red, last 2 panels) and confocal microscopy analysis. Original magnification $\times 63$. (D) Dot plots showing Annexin-FITC (AV) and propidium iodide (PI) staining of CD19⁺ cells cultured with NLC derived with or without NAMPT for 14 days. The graph represents cumulative data ($n = 14$). (E) CFSE-labeled, preactivated (anti-CD3/IL-2, 24 hours) autologous PBMCs were cocultured with predifferentiated NLCs (3-5 days with or without rNAMPT). Graph shows cumulative data ($n = 8$). (F) FACS analysis of basal Treg (CD4⁺/CD25^{high}/CD127^{low}) expression and after 5 days of coculture (with or without rNAMPT) with autologous NLCs. The gray histogram represents the isotype control. Box plot represent the cumulative data ($n = 9$) showing the percentage of PD-1 expression. (G) FACS analysis of PD-1 expression on CD4⁺ T cells after 5 days of coculture (with or without rNAMPT) with autologous NLCs. The gray histogram represents the isotype control. Box plot represent the cumulative data ($n = 9$) showing the percentage of PD-1 expression. (H) PD-L1 expression on NLCs treated with rNAMPT for 24 hours was checked by flow cytometry and confirmed by quantitative PCR ($n = 14$).

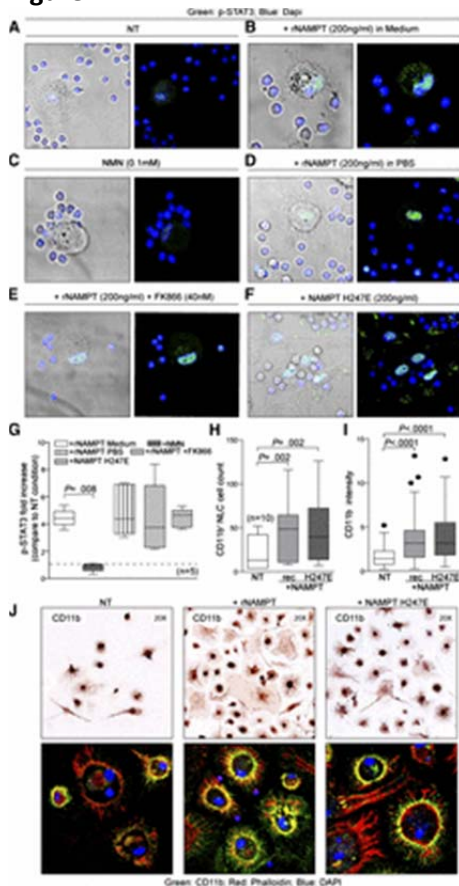
NLC generated in the presence of rNAMPT had a greater ability to sustain leukemic cells (Figure 6D). Conversely, when autologous T lymphocytes preactivated using anti-CD3 antibody and IL-2 were cultured with NLCs in the presence of rNAMPT, proliferation was markedly suppressed (Figure 6E). These culture conditions also increased the number of regulatory T cells (Treg's), as determined on the basis of a CD4⁺/CD25^{high}/CD127^{low} phenotype (Figure 6F). Lastly, culture of T cells in the presence of rNAMPT-treated NLCs led to the upmodulation of the PD-1/PD-L1 axis (Figure 6G-H).⁶⁶ Upon total PBMCs coculturing with autologous NLCs in the presence of rNAMPT, CD4⁺ T lymphocytes showed higher PD-1 expression than when the same cells were cultured with NLCs without rNAMPT (Figure 6G). Consistently, in the presence of rNAMPT, NLCs expressed higher levels of PD-L1 (Figure 6H).

Together, these findings suggest that eNAMPT is among the soluble factors produced by leukemic cells to turn nonneoplastic bystander cells into supporters of tumor growth and survival.

eNAMPT effects are independent of the enzymatic functions of the molecule

To explore the role of enzymatic activity in modulating eNAMPT effects, we selected STAT3 activation and NLC differentiation as prototypes of short- and long-term rNAMPT effects. Four lines of evidence argue in favor of a nonenzymatic function of eNAMPT in this model. First, exposure of NLCs to extracellular NMN failed to induce STAT3 phosphorylation, suggesting that the product of the reaction is inactive (Figure 7A-C). Second, treatment of NLCs with rNAMPT diluted in PBS, without medium that contains high levels of nicotinamide, the substrate of the reaction, induced prominent activation of STAT3 (Figure 7D). Third, treatment with the NAMPT inhibitor FK866 did not block the ability of rNAMPT to stimulate the phosphorylation of STAT3 (Figure 7E). Fourth, the enzymatically inactive NAMPT H247E mutant⁶⁷ retained the ability to activate STAT3 phosphorylation (Figure 7F-G).²² Considered together, these results indicate that the enzymatic activity of eNAMPT is dispensable when triggering NLC activation. Similarly, the H247E mutant enhanced NLC differentiation, as determined by morphologic parameters and protein expression (CD11b and CD68; Figure 7H-J and supplemental Figure 12A-B).

Figure 7



eNAMPT activities are independent of the enzymatic activities. (A-F) NLCs were treated (30 minutes) in different conditions and then stained for the presence of p-STAT3 (green fluorescence). DAPI (blue) was used to visualize nuclei. (G) Cumulative data ($n = 5$) are shown. (H) Box plots representing the quantification of CD11b⁺ NLCs count ($n = 10$) and the corresponding fluorescence pixel intensity (I) in the indicated conditions. (J) CD11b staining in NLC preparation by immunocytochemistry (upper panels, original magnification ×20) and by immunofluorescence (green, bottom panels, original magnification ×63).

Overall these results indicate that the enzymatic activity of eNAMPT is not essential for activating signaling or inducing long-term effects such as NLC differentiation.

Discussion

This study demonstrates that NAMPT possesses 2 distinct functions in the CLL microenvironment. Inside the cell, it regulates production of NMN from nicotinamide, the limiting step in the generation of NAD. CLL lymphocytes expressed higher levels of iNAMPT and contained higher amounts of NMN and of NAD than

circulating normal B lymphocytes from donors of similar ages, a finding consistent with increased NAD needs to allow for continued glycolytic flux.⁶⁸ Consistently, iNAMPT levels were further upregulated when CLL cells were activated through distinct receptors, suggesting also that *NAMPT* transcription may be controlled by signaling intermediates common to these pathways, including BTK. The significant difference in NAD levels between CLL cells and normal B lymphocytes was also noted in a previous study using a different method.⁶⁹ These data are also in line with reports indicating that CLL lymphocytes are particularly sensitive to the NAMPT inhibitor FK866, which induces rapid NAD deprivation.⁷⁰⁻⁷²

The second main finding of this work is that elevated amounts of eNAMPT are present in the plasma of CLL patients. In the CLL samples examined, eNAMPT levels were significantly higher than those scored by controls, which were in line with the literature.⁵⁸ After correcting for the number of circulating monocytes, we observed a direct correlation between eNAMPT levels and lymphocytosis, indirectly suggesting that the leukemic cells contribute to eNAMPT production. In agreement with this hypothesis, activation of purified CLL lymphocytes increased the concentrations of eNAMPT, and within LN proliferating CLL cells were clearly NAMPT⁺.

eNAMPT produced by CLL cells enhanced polarization of circulating monocytes into macrophages with an M2 phenotype. This conclusion was reached after showing (1) expression of nuclear factors and surface markers involved in M2 differentiation, (2) secretion of immunomodulatory cytokines/chemokines, and (3) production of mediators of immune suppression. The use of a blocking anti-NAMPT antibody significantly compromised NLC differentiation and phenotype, arguing for a direct role of the molecule in the acquisition of M2 properties.

NAMPT is also part of the M2 signature, as inferred from the finding that both monocytes and NLCs contained high levels of iNAMPT and could produce eNAMPT. Consistently, the use of lenalidomide to correct M2 polarization of NLCs was followed by a marked decrease of iNAMPT and eNAMPT.

When NLCs were exposed to NAMPT ERK1/2, STAT3 and NF- κ B were activated, arguing for a direct effect of this protein on the target cells. The blocking anti-NAMPT antibody completely abrogated pathway activation, in line with a receptor-mediated effect. Culture with rNAMPT also enhanced the immunosuppressive phenotype of NLCs, as well as their ability to sustain CLL survival and suppress autologous T-cell proliferation and Treg expansion. This could be achieved through the production of IL-6 and IL-8 as tumor-promoting cytokines,^{73,74} while IL-10, CCL18, and IDO could act as immunosuppressors.⁷⁵

One of the intriguing questions in the field concerns the role of the enzymatic activity in the functions exerted by eNAMPT. Our data indicated that eNAMPT is catalytically active, as expected,^{76,77} even if NMN, the reaction product, was undetectable both in plasma and in media, confirming previous data.⁷⁸ This finding, together with the observation that eNAMPT might be unable to catalyze NMN formation in the extracellular space because of the lack of suitable substrate concentrations (particularly PRPP),⁵³ suggests that the functional responses elicited by eNAMPT are independent of its catalytic activity. Four independent lines of evidence, including the use of inactive eNAMPT mutant (H247E),⁶⁷ are provided to support the conclusion that NAMPT activity is dispensable to activate STAT3 and NF- κ B pathways in NLCs and long-term NLC differentiation. This finding leaves open the issue of how eNAMPT may exert its functional activities. Future studies will tell whether there is a NAMPT receptor or whether alternative mechanisms, such as internalization of the enzyme, are in place.

From the translational point of view, an important observation is that drugs interfering with CLL signaling, such as ibrutinib, can suppress *NAMPT* transcription, reducing both iNAMPT and eNAMPT. Furthermore, NLCs differentiated in the presence of lenalidomide express lower levels of NAMPT, suggesting that drugs that restore immune functions interfere with the production of eNAMPT.

To conclude, we propose that a vicious circle based on CLL cell activation through antigen and accessory signals increases eNAMPT and CCL3 production. CCL3 serves as an attractant for circulating monocytes, which, in the presence of high levels of eNAMPT, differentiate into NLCs, with an enhanced M2 phenotype and functional characteristics, contributing to CLL survival, activation, and proliferation and inhibition of T-cell responses.

References

1. Sica A, Allavena P, Mantovani A. Cancer related inflammation: the macrophage connection. *Cancer Lett* 2008;267(2):204-215.
2. Qian BZ, Pollard JW. Macrophage diversity enhances tumor progression and metastasis. *Cell* 2010;141(1):39-51.
3. Biswas SK, Mantovani A. Macrophage plasticity and interaction with lymphocyte subsets: cancer as a paradigm. *Nat Immunol* 2010;11(10):889-896.
4. Coussens LM, Werb Z. Inflammation and cancer. *Nature* 2002;420(6917):860-867.
5. Burger JA, Ghia P, Rosenwald A, Caligaris-Cappio F. The microenvironment in mature B-cell malignancies: a target for new treatment strategies. *Blood* 2009;114(16):3367-3375.
6. Zenz T, Mertens D, Küppers R, Döhner H, Stilgenbauer S. From pathogenesis to treatment of chronic lymphocytic leukaemia. *Nat Rev Cancer* 2010;10(1):37-50.
7. Audrito V, Vaisitti T, Serra S, et al. Targeting the microenvironment in chronic lymphocytic leukemia offers novel therapeutic options. *Cancer Lett* 2013;328(1):27-35.
8. Ponzoni M, Doglioni C, Caligaris-Cappio F. Chronic lymphocytic leukemia: the pathologist's view of lymph node microenvironment. *Semin Diagn Pathol* 2011;28(2):161-166.
9. Fecteau JF, Kipps TJ. Structure and function of the hematopoietic cancer niche: focus on chronic lymphocytic leukemia. *Front Biosci (Schol Ed)* 2012;4:61-73.
10. Ferretti E, Bertolotto M, Deaglio S, et al. A novel role of the CX3CR1/CX3CL1 system in the cross-talk between chronic lymphocytic leukemia cells and tumor microenvironment. *Leukemia* 2011;25(8):1268-1277.
11. Ysebaert L, Fournié JJ. Genomic and phenotypic characterization of nurse-like cells that promote drug resistance in chronic lymphocytic leukemia. *Leuk Lymphoma* 2011;52(7):1404-1406.
12. Filip AA, Ciseł B, Koczkodaj D, Wąsik-Szczepanek E, Piersiak T, Dmoszyńska A. Circulating microenvironment of CLL: are nurse-like cells related to tumor-associated macrophages? *Blood Cells Mol Dis* 2013;50(4):263-270.
13. Maffei R, Bulgarelli J, Fiorcari S, et al. The monocytic population in chronic lymphocytic leukemia shows altered composition and deregulation of genes involved in phagocytosis and inflammation. *Haematologica* 2013;98(7):1115-1123.
14. Burger JA, Tsukada N, Burger M, Zvaifler NJ, Dell'Aquila M, Kipps TJ. Blood-derived nurse-like cells protect chronic lymphocytic leukemia B cells from spontaneous apoptosis through stromal cell-derived factor-1. *Blood* 2000;96(8):2655-2663.
15. Burger JA, Kipps TJ. Chemokine receptors and stromal cells in the homing and homeostasis of chronic lymphocytic leukemia B cells. *Leuk Lymphoma* 2002;43(3):461-466.
16. Tsukada N, Burger JA, Zvaifler NJ, Kipps TJ. Distinctive features of "nurselike" cells that differentiate in the context of chronic lymphocytic leukemia. *Blood* 2002;99(3):1030-1037.
17. Samal B, Sun Y, Stearns G, Xie C, Suggs S, McNiece I. Cloning and characterization of the cDNA encoding a novel human pre-B-cell colony-enhancing factor. *Mol Cell Biol* 1994;14(2):1431-1437.

18. Chang YH, Chang DM, Lin KC, Shin SJ, Lee YJ. Visfatin in overweight/obesity, type 2 diabetes mellitus, insulin resistance, metabolic syndrome and cardiovascular diseases: a meta-analysis and systemic review. *Diabetes Metab Res Rev* 2011;27(6):515-527.
19. Martin PR, Shea RJ, Mulks MH. Identification of a plasmid-encoded gene from *Haemophilus ducreyi* which confers NAD independence. *J Bacteriol* 2001;183(4):1168-1174.
20. Rongvaux A, Shea RJ, Mulks MH, et al. Pre-B-cell colony-enhancing factor, whose expression is up-regulated in activated lymphocytes, is a nicotinamide phosphoribosyltransferase, a cytosolic enzyme involved in NAD biosynthesis. *Eur J Immunol* 2002;32(11):3225-3234.
21. Houtkooper RH, Cantó C, Wanders RJ, Auwerx J. The secret life of NAD⁺: an old metabolite controlling new metabolic signaling pathways. *Endocr Rev* 2010;31(2):194-223.
22. Li Y, Zhang Y, Dorweiler B, et al. Extracellular Nampt promotes macrophage survival via a nonenzymatic interleukin-6/STAT3 signaling mechanism. *J Biol Chem* 2008;283(50):34833-34843.
23. Liu P, Li H, Cepeda J, et al. Regulation of inflammatory cytokine expression in pulmonary epithelial cells by pre-B-cell colony-enhancing factor via a nonenzymatic and AP-1-dependent mechanism. *J Biol Chem* 2009;284(40):27344-27351.
24. Sun Z, Lei H, Zhang Z. Pre-B cell colony enhancing factor (PBEF), a cytokine with multiple physiological functions. *Cytokine Growth Factor Rev* 2013;24(5):433-442.
25. Zhang LQ, Heruth DP, Ye SQ. Nicotinamide phosphoribosyltransferase in human diseases. *J Bioanal Biomed* 2011;3:13-25.
26. Gallí M, Van Gool F, Rongvaux A, Andris F, Leo O. The nicotinamide phosphoribosyltransferase: a molecular link between metabolism, inflammation, and cancer. *Cancer Res* 2010;70(1):8-11.
27. Garten A, Petzold S, Körner A, Imai S, Kiess W. Nampt: linking NAD biology, metabolism and cancer. *Trends Endocrinol Metab* 2009;20(3):130-138.
28. Imai S. Nicotinamide phosphoribosyltransferase (Nampt): a link between NAD biology, metabolism, and diseases. *Curr Pharm Des* 2009;15(1):20-28.
29. Jieyu H, Chao T, Mengjun L, et al. Nampt/visfatin/PBEF: a functionally multi-faceted protein with a pivotal role in malignant tumors. *Curr Pharm Des* 2012;18(37):6123-6132.
30. Vaisitti T, Audrito V, Serra S, et al. The enzymatic activities of CD38 enhance CLL growth and trafficking: implications for therapeutic targeting [published online ahead of print July 3, 2014]. *Leukemia*.
31. Deaglio S, Vaisitti T, Bergui L, et al. CD38 and CD100 lead a network of surface receptors relaying positive signals for B-CLL growth and survival. *Blood* 2005;105(8):3042-3050.
32. Hong SB, Huang Y, Moreno-Vinasco L, et al. Essential role of pre-B-cell colony enhancing factor in ventilator-induced lung injury. *Am J Respir Crit Care Med* 2008;178(6):605-617.
33. Pillai VB, Sundaresan NR, Kim G, et al. Nampt secreted from cardiomyocytes promotes development of cardiac hypertrophy and adverse ventricular remodeling. *Am J Physiol Heart Circ Physiol* 2013;304(3):H415-H426.
34. Zamporlini F, Ruggieri S, Mazzola F, Amici A, Orsomando G, Raffaelli N. Novel assay for simultaneous measurement of pyridine mononucleotides synthesizing activities allows dissection of the NAD(+) biosynthetic machinery in mammalian cells [published online ahead of print October 4, 2014]. *FEBS J*.
35. Graeff R, Lee HC. A novel cycling assay for nicotinic acid-adenine dinucleotide phosphate with nanomolar sensitivity. *Biochem J* 2002;367(pt 1):163-168.
36. Formentini L, Moroni F, Chiarugi A. Detection and pharmacological modulation of nicotinamide mononucleotide (NMN) in vitro and in vivo. *Biochem Pharmacol* 2009;77(10):1612-1620.

37. Deaglio S, Capobianco A, Bergui L, et al. CD38 is a signaling molecule in B-cell chronic lymphocytic leukemia cells. *Blood* 2003;102(6):2146-2155.
38. Serra S, Horenstein AL, Vaisitti T, et al. CD73-generated extracellular adenosine in chronic lymphocytic leukemia creates local conditions counteracting drug-induced cell death. *Blood* 2011;118(23):6141-6152.
39. Tsai YM, Hsu SC, Zhang J, et al. Functional interaction of cockroach allergens and mannose receptor (CD206) in human circulating fibrocytes. *PLoS ONE* 2013;8(5):e64105.
40. Arruga F, Gizdic B, Serra S, et al. Functional impact of NOTCH1 mutations in chronic lymphocytic leukemia. *Leukemia* 2014;28(5):1060-1070.
41. Vaisitti T, Aydin S, Rossi D, et al. CD38 increases CXCL12-mediated signals and homing of chronic lymphocytic leukemia cells. *Leukemia* 2010;24(5):958-969.
42. Audrito V, Vaisitti T, Rossi D, et al. Nicotinamide blocks proliferation and induces apoptosis of chronic lymphocytic leukemia cells through activation of the p53/miR-34a/SIRT1 tumor suppressor network. *Cancer Res* 2011;71(13):4473-4483.
43. Giannoni P, Pietra G, Travaini G, et al. Chronic lymphocytic leukemia nurse-like cells express hepatocyte growth factor receptor (c-MET) and indoleamine 2,3-dioxygenase and display features of immunosuppressive type 2 skewed macrophages. *Haematologica* 2014;99(6):1078-1087.
44. Revollo JR, Grimm AA, Imai S. The NAD biosynthesis pathway mediated by nicotinamide phosphoribosyltransferase regulates Sir2 activity in mammalian cells. *J Biol Chem* 2004;279(49):50754-50763.
45. Zucchetto A, Benedetti D, Tripodo C, et al. CD38/CD31, the CCL3 and CCL4 chemokines, and CD49d/vascular cell adhesion molecule-1 are interchained by sequential events sustaining chronic lymphocytic leukemia cell survival. *Cancer Res* 2009;69(9):4001-4009.
46. Guarini A, Chiaretti S, Tavolaro S, et al. BCR ligation induced by IgM stimulation results in gene expression and functional changes only in IgV H unmutated chronic lymphocytic leukemia (CLL) cells. *Blood* 2008;112(3):782-792.
47. Kenny EF, Quinn SR, Doyle SL, Vink PM, van Eenennaam H, O'Neill LA. Bruton's tyrosine kinase mediates the synergistic signalling between TLR9 and the B cell receptor by regulating calcium and calmodulin. *PLoS ONE* 2013;8(8):e74103.
48. Rodríguez-Alba JC, Moreno-García ME, Sandoval-Montes C, Rosales-Garcia VH, Santos-Argumedo L. CD38 induces differentiation of immature transitional 2 B lymphocytes in the spleen. *Blood* 2008;111(7):3644-3652.
49. Herman SE, Gordon AL, Hertlein E, et al. Bruton tyrosine kinase represents a promising therapeutic target for treatment of chronic lymphocytic leukemia and is effectively targeted by PCI-32765. *Blood* 2011;117(23):6287-6296.
50. Friebe D, Neef M, Kratzsch J, et al. Leucocytes are a major source of circulating nicotinamide phosphoribosyltransferase (NAMPT)/pre-B cell colony (PBEF)/visfatin linking obesity and inflammation in humans. *Diabetologia* 2011;54(5):1200-1211.
51. Jacques C, Holzenberger M, Mladenovic Z, et al. Proinflammatory actions of visfatin/nicotinamide phosphoribosyltransferase (Nampt) involve regulation of insulin signaling pathway and Nampt enzymatic activity. *J Biol Chem* 2012;287(18):15100-15108.
52. Romacho T, Azcutia V, Vázquez-Bella M, et al. Extracellular PBEF/NAMPT/visfatin activates pro-inflammatory signalling in human vascular smooth muscle cells through nicotinamide phosphoribosyltransferase activity. *Diabetologia* 2009;52(11):2455-2463.

53. Hara N, Yamada K, Shibata T, Osago H, Tsuchiya M. Nicotinamide phosphoribosyltransferase/visfatin does not catalyze nicotinamide mononucleotide formation in blood plasma. *PLoS ONE* 2011;6(8):e22781.
54. Cols M, Barra CM, He B, et al. Stromal endothelial cells establish a bidirectional crosstalk with chronic lymphocytic leukemia cells through the TNF-related factors BAFF, APRIL, and CD40L. *J Immunol* 2012;188(12):6071-6083.
55. Ferrer G, Bosch R, Hodgson K, et al. B cell activation through CD40 and IL4R ligation modulates the response of chronic lymphocytic leukaemia cells to BAFF and APRIL. *Br J Haematol* 2014;164(4):570-578.
56. Yun MR, Seo JM, Park HY. Visfatin contributes to the differentiation of monocytes into macrophages through the differential regulation of inflammatory cytokines in THP-1 cells. *Cell Signal* 2014;26(4):705-715.
57. Skokowa J, Lan D, Thakur BK, et al. NAMPT is essential for the G-CSF-induced myeloid differentiation via a NAD(+)-sirtuin-1-dependent pathway. *Nat Med* 2009;15(2):151-158.
58. Moschen AR, Kaser A, Enrich B, et al. Visfatin, an adipocytokine with proinflammatory and immunomodulating properties. *J Immunol* 2007;178(3):1748-1758.
59. Zucchetto A, Tripodo C, Benedetti D, et al. Monocytes/macrophages but not T lymphocytes are the major targets of the CCL3/CCL4 chemokines produced by CD38(+)CD49d(+) chronic lymphocytic leukaemia cells. *Br J Haematol* 2010;150(1):111-113.
60. Ko SY, Ladanyi A, Lengyel E, Naora H. Expression of the homeobox gene HOXA9 in ovarian cancer induces peritoneal macrophages to acquire an M2 tumor-promoting phenotype. *Am J Pathol* 2014;184(1):271-281.
61. Martinez FO, Gordon S, Locati M, Mantovani A. Transcriptional profiling of the human monocyte-to-macrophage differentiation and polarization: new molecules and patterns of gene expression. *J Immunol* 2006;177(10):7303-7311.
62. Jia L, Clear A, Liu FT, et al. Extracellular HMGB1 promotes differentiation of nurse-like cells in chronic lymphocytic leukemia. *Blood* 2014;123(11):1709-1719.
63. Fiorcari S, Martinelli S, Bulgarelli J, et al. Lenalidomide interferes with tumor-promoting properties of nurse-like cells in chronic lymphocytic leukemia. *Haematologica*. In press.
64. Moros A, Rodríguez V, Saborit-Villarroya I, et al. Synergistic antitumor activity of lenalidomide with the BET bromodomain inhibitor CPI203 in bortezomib-resistant mantle cell lymphoma. *Leukemia* 2014;28(10):2049-2059.
65. Fan Y, Meng S, Wang Y, Cao J, Wang C. Visfatin/PBEF/Nampt induces EMMPRIN and MMP-9 production in macrophages via the NAMPT-MAPK (p38, ERK1/2)-NF- κ B signaling pathway. *Int J Mol Med* 2011;27(4):607-615.
66. Brusa D, Serra S, Coscia M, et al. The PD-1/PD-L1 axis contributes to T-cell dysfunction in chronic lymphocytic leukemia. *Haematologica* 2013;98(6):953-963.
67. Wang T, Zhang X, Bheda P, Revollo JR, Imai S, Wolberger C. Structure of Nampt/PBEF/visfatin, a mammalian NAD⁺ biosynthetic enzyme. *Nat Struct Mol Biol* 2006;13(7):661-662.
68. Chiarugi A, Dölle C, Felici R, Ziegler M. The NAD metabolome—a key determinant of cancer cell biology. *Nat Rev Cancer* 2012;12(11):741-752.
69. Liebes LF, Krigel RL, Conklyn M, Nevrla DR, Silber R. Ribonucleotide content of mononuclear cells from normal subjects and patients with chronic lymphocytic leukemia: increased nicotinamide adenine dinucleotide concentration in chronic lymphocytic leukemia lymphocytes. *Cancer Res* 1983;43(11):5608-5617.

70. Nahimana A, Attinger A, Aubry D, et al. The NAD biosynthesis inhibitor APO866 has potent antitumor activity against hematologic malignancies. *Blood* 2009;113(14):3276-3286.
71. Montecucco F, Cea M, Bauer I, et al. Nicotinamide phosphoribosyltransferase (NAMPT) inhibitors as therapeutics: rationales, controversies, clinical experience. *Curr Drug Targets* 2013;14(6):637-643.
72. Gehrke I, Bouchard ED, Beiggi S, et al. On-target effect of FK866, a nicotinamide phosphoribosyl transferase inhibitor, by apoptosis-mediated death in chronic lymphocytic leukemia cells. *Clin Cancer Res* 2014;20(18):4861-4872.
73. Schafer ZT, Brugge JS. IL-6 involvement in epithelial cancers. *J Clin Invest* 2007;117(12):3660-3663.
74. Xu H, Lai W, Zhang Y, et al. Tumor-associated macrophage-derived IL-6 and IL-8 enhance invasive activity of LoVo cells induced by PRL-3 in a KCNN4 channel-dependent manner. *BMC Cancer* 2014;14(1):330.
75. Uyttenhove C, Pilotte L, Théate I, et al. Evidence for a tumoral immune resistance mechanism based on tryptophan degradation by indoleamine 2,3-dioxygenase. *Nat Med* 2003;9(10):1269-1274.
76. Revollo JR, Körner A, Mills KF, et al. Nampt/PBEF/visfatin regulates insulin secretion in beta cells as a systemic NAD biosynthetic enzyme. *Cell Metab* 2007;6(5):363-375.
77. Kover K, Tong PY, Watkins D, et al. Expression and regulation of Nampt in human islets. *PLoS ONE* 2013;8(3):e58767.
78. Yamada K, Hara N, Shibata T, Osago H, Tsuchiya M. The simultaneous measurement of nicotinamide adenine dinucleotide and related compounds by liquid chromatography/electrospray ionization tandem mass spectrometry. *Anal Biochem* 2006;352(2):282-285.